

**Protective effect of  $\gamma$ -mangostin isolated from the peel of *Garcinia mangostana* against glutamate-induced cytotoxicity in HT22 hippocampal neuronal cells**

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## Extraction and isolation

The dried peel of *Garcinia mangostana* L (1.23kg) was extracted with EtOH three times. The EtOH extract (87.1g) was suspended in water and then partitioned with CHCl<sub>3</sub>, EtOAc and *n*-BuOH, yielding 50.18g, 2.74g, and 18.27g, respectively. The CHCl<sub>3</sub> fraction (45.93g) was chromatographed over a silica gel column chromatography using gradient with CHCl<sub>3</sub>- MeOH (100:0 to 0:100) as solvent and obtained 18 subfraction (GMC1-GMC18). GMC12 (4.73 g) fraction was separated using preparative RP-MPLC with gradient mixture of 40% to 80% MeOH to yield  $\gamma$ -mangostin (3.20 g). The structure of  $\gamma$ -mangostin was confirmed by comparing the measured NMR data with the published values [1,2].

**$\gamma$ -mangostin:** HRESIMS:  $m/z$  397.1652 [M+H]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>25</sub>O<sub>6</sub>, 397.1651) <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz)  $\delta$  6.67 (1H, s, H-5), 6.24 (1H, s, H-4), 5.25 (2H, m, H-2', 2''), 4.12 (2H, d,  $J$  = 5.9 Hz, H-1''), 3.34 (2H, m, H-1'), 1.84 (3H, s, H-4''), 1.78 (3H, s, H-4'), 1.66 (6H, s, H-5', 5''); <sup>13</sup>CNMR (CD<sub>3</sub>OD, 100 MHz)  $\delta$  183.8 (C-9), 163.5 (C-3), 161.7 (C-1), 156.5 (C-6), 154.2 (C-4a), 153.4 (C-10a), 142.2 (C-7), 131.9 (C-3'), 131.8 (C-3''), 129.8 (C-8), 125.0 (C-2''), 124.1 (C-2'), 112.4 (C-2), 111.3 (C-8a), 104.1 (C-9a), 101.1 (C-5), 93.1 (C-4), 26.8 (C-1''), 26.2 (C-5''), 26.1 (C-5'), 22.4 (C-1'), 18.5 (C-4'), 18.1 (C-4'').

## UPLC-UV analysis for the purity of $\gamma$ -mangostin

Chromatographic analysis for the purity of  $\gamma$ -mangostin was performed on an Acquity UPLC system with an ACQUITY UPLC®BEH C18 column (2.1 ×150 mm, 1.7  $\mu$ m, Waters, Milford, MA, USA). An aliquot (3  $\mu$ L) of the [sample \(0.1mg/mL in MeOH\)](#) was injected into the UPLC system. The mobile phase consisted of 0.1% formic acid in water (A) and 0.1% formic acid in acetonitrile (B). The mobile phase consisting of (A) and (B) was run at a flow rate of 0.3 mL/min by the following programmed gradient elution: 40% (B, v/v) isocratic for 15min, 40→100% (B) in 5min, 100% (B) isocratic for 3min, 100→40% (B) in 0.5min, 40% (B) isocratic for 1.5min as post-run for reconditioning. The column temperature was maintained at 40 °C and the wavelength was set at 320 nm

## Reference

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2. Chin, Xiao-Qian; Zi, Cheng-Ting; Li, Hong-Mei; Yang, Liu; Lv, Yong-Feng; Li, Jin-Yu; Hpu, Bo; Ren, Fu-Cai; Hu, Jiang-Miao; Zhou, Jun. Design, synthesis and structure-activity relationships of mangostin analogs as cytotoxic agents. *RSC Advances*, 2018, 8, 72, 41377-41388.

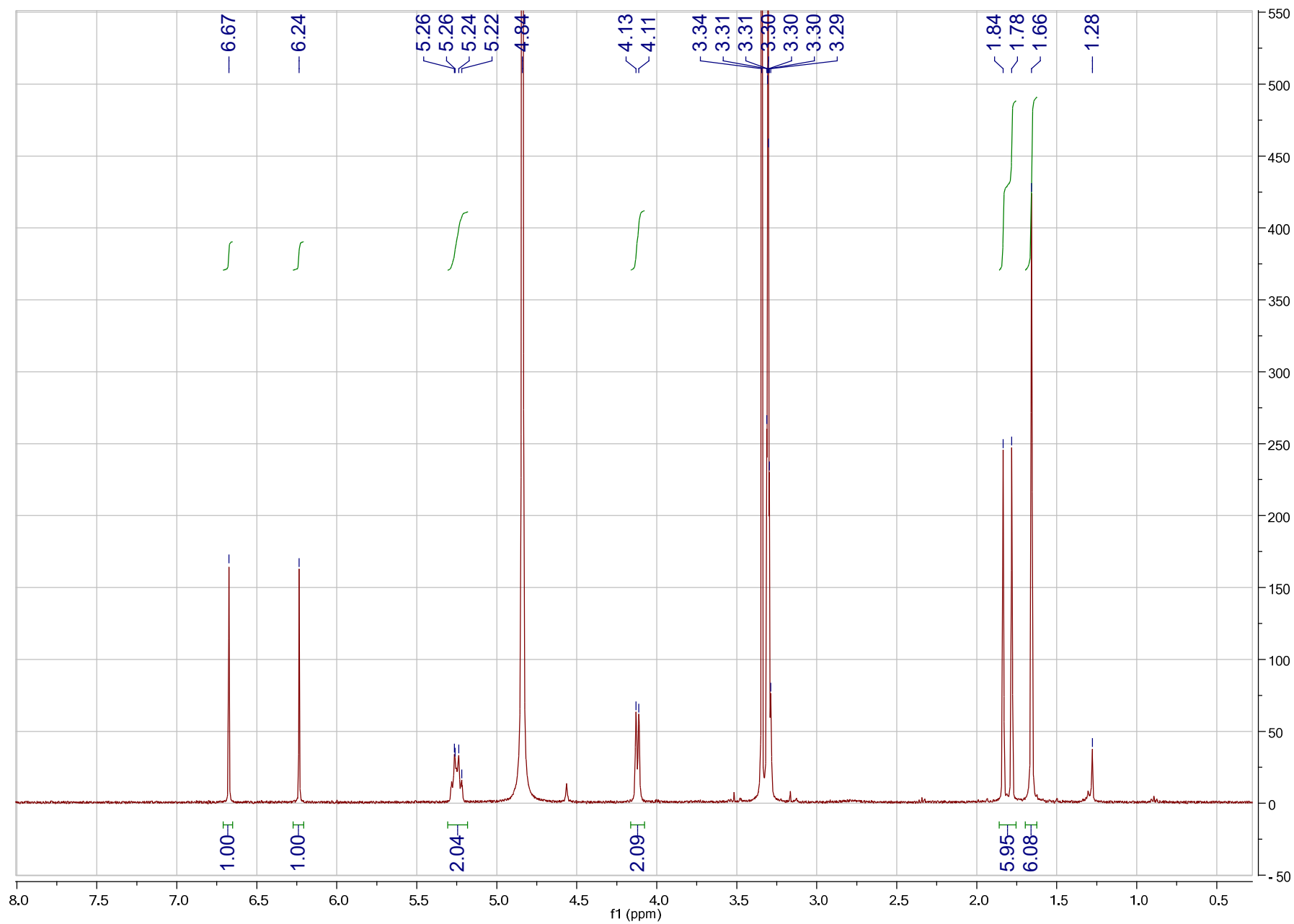


Figure S1.  $^1\text{H}$  NMR spectrum of  $\gamma$ -mangostin in  $\text{CD}_3\text{OD}$ , 400 MHz

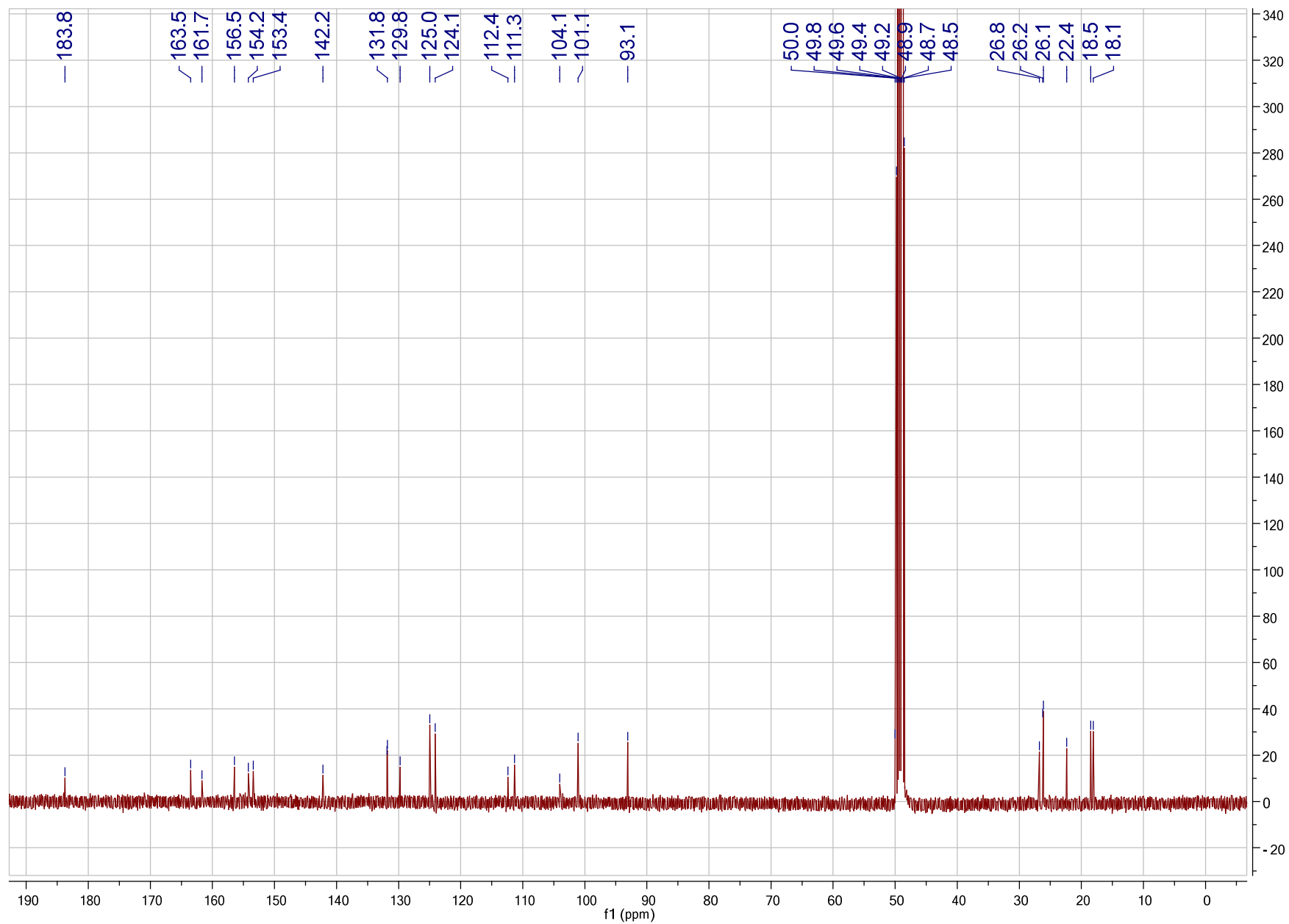


Figure S2. <sup>13</sup>C NMR spectrum of  $\gamma$ -mangostin ( $\text{CD}_3\text{OD}$ , 100 MHz)

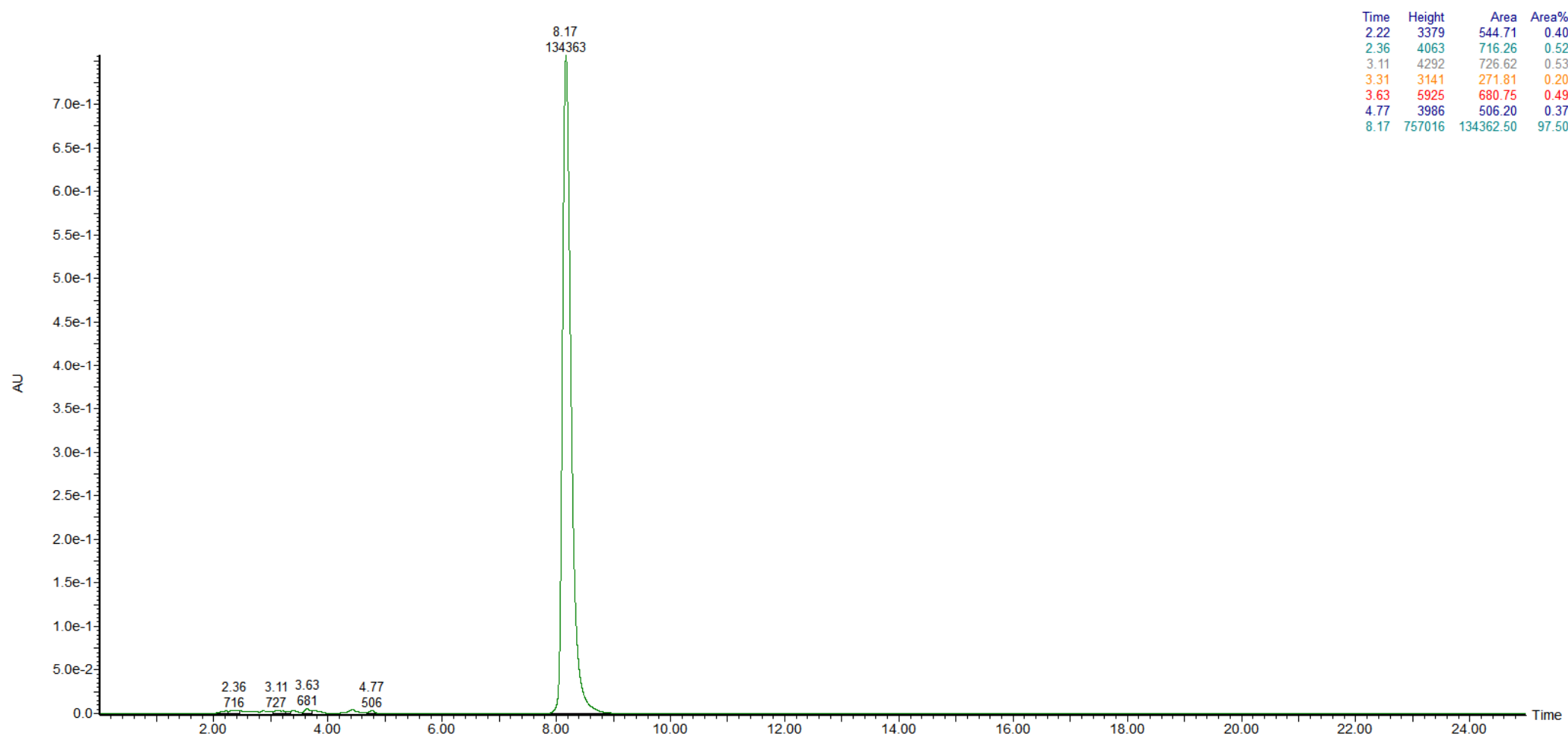


Figure S3. The UPLC UV chromatogram of  $\gamma$ -mangostin (UV wavelength 320nm)

gamma-MS5 558 (9.574)

1: TOF MS ES+  
1.56e4

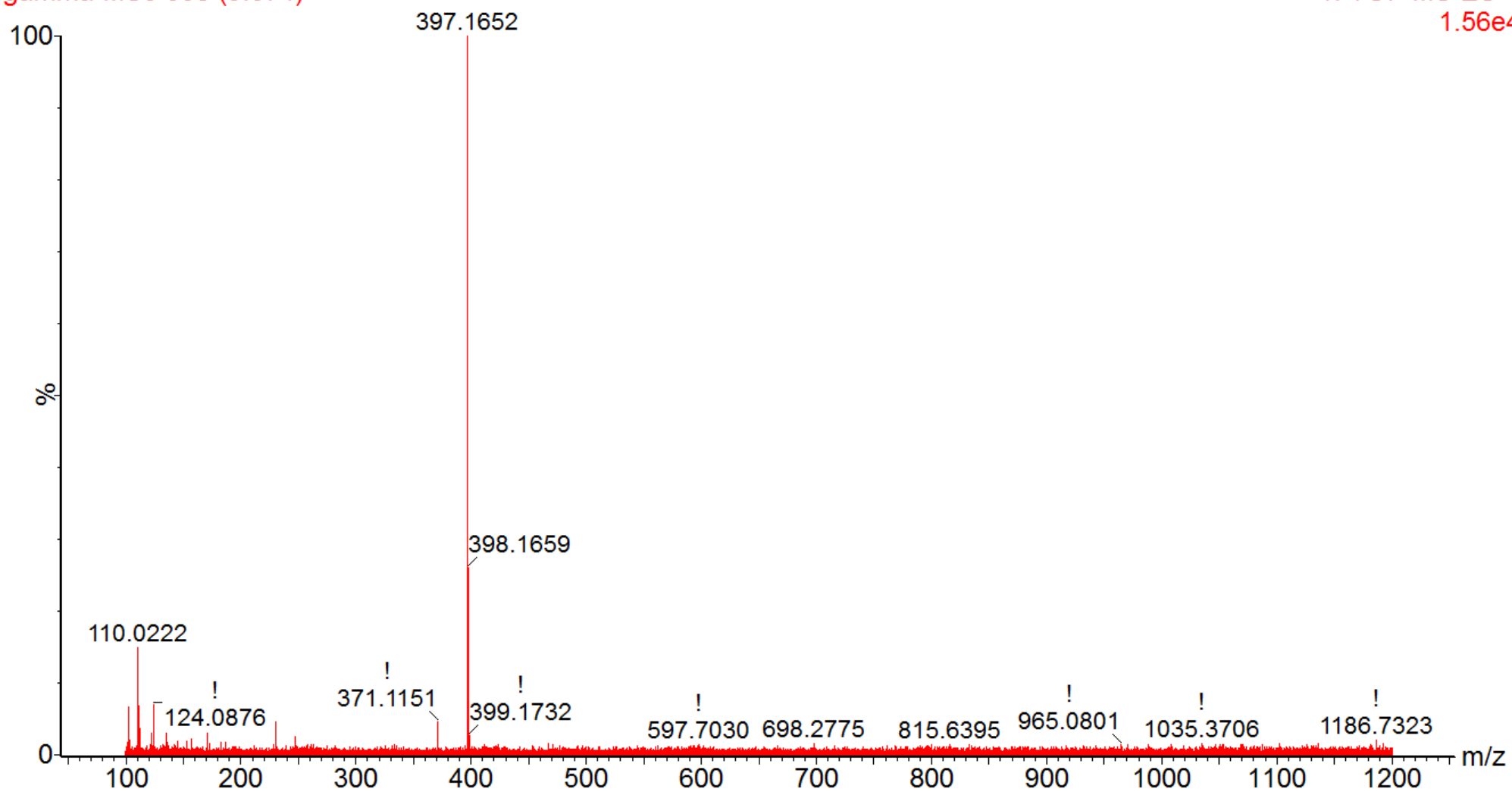


Figure S4. The MASS spectral data of  $\gamma$ -mangostin